REVIEW ARTICLE

Epigenetic mechanisms in endometrial cancer

Albiona Stampoliou¹, Petroula Arapantoni-Dadioti², Kitty Pavlakis³

¹MSc Programme «Research on Female Reproduction» Athens University Medical School & Midwifery Department, T.E.I. of Athens, Athens; ²Pathology Department, «Metaxa» Cancer Hospital, Piraeus; ³Pathology Department, Athens University Medical School, Athens, Greece

Summary

Purpose: Endometrial cancer is a very common type of cancer in females worldwide. Advances in diagnosis and treatment have not decreased the incidence of endometrial cancer. Lately, research has been focused on revealing the molecular and genetic characteristics of endometrial cancer in order to provide new insights in the biology of this entity, leading hopefully to innovating therapies. Research has revealed that epigenetic modifications govern endometrial

carcinogenesis. In this review, the epigenetic mechanisms that are involved in endometrial cancer as well as the differences between the different types of endometrial cancer are discussed. The review also refers to the putative therapeutic benefits that hopefully can arise.

Key words: endometrial cancer, endometrioid, epigenetic, methylation

Introduction

Endometrial cancer is the fourth most common malignancy in women in Europe and the most common gynecologic malignancy in the United States [1]. The incidence of endometrial cancer has increased in the last years and despite advances in diagnosis and treatment, the death rates have steadily been increasing over the past 20 years [2]. Recent progression in research has revealed extensive epigenetic modifications that are involved in endometrial carcinogenesis and offer a window of opportunity in improved therapies. In this review, the epigenetic mechanisms involved in endometrial cancer are discussed, as well as the putative therapeutic benefits that can arise.

Epigenetics

According to the NIH "Roadmap Epigenomics Project," the term epigenetics refers to both heritable changes in gene activity and expression (in the progeny of cells or of individuals) and also stable, long-term alterations in the transcriptional potential of a cell that are not necessarily heritable. Epigenetic modifications are defined as any modifications in genomic DNA that do not allow transcription of DNA thus causing transcriptional silencing. These modifications are not affected by cell division, they do not alter the genome's sequence and they can be both beneficial and detrimental. In cancer, for example, epigenetic modifications keep the genome safe by not allowing rearrangements in chromatin that can cause high gene activation but can also be harmful through silencing of tumor suppressor genes. Comprehension of the epigenetic mechanisms in carcinogenesis is valuable for developing and ameliorating cancer treatment and prevention [3]. There are three different types of mechanisms that cause gene silencing: DNA methylation, histone modifications and RNA-associated silencing.

DNA methylation is now considered a hall-

Correspondence to: Albiona Stampoliou, MSc. Eleftheriou Venizelou 79, 18863, Perama, Athens, Greece. Tel: +30 6976267187, E-mail: salbiona@gmail.com

Received: 05/10/2015; Accepted: 18/10/2015

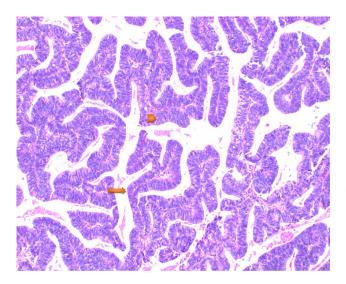


Figure 1. Well differentiated endometrioid adenocarcinoma (H&E x400). Glandular cells with nuclear stratification featuring minimal to moderate atypia (arrowheads). There is a sharp demarcation of the apical border of the neoplastic glands (arrows).

mark of cancer. DNA is methylated by DNA cytosine methyltransferases (DNMT1, 3A and 3B), that catalyze the transfer of a methyl group to a cytosine nucleotide next to a guanine (CpG) and 5-methyl-2'-deoxycytidine (5-mC) is formed. These CpG clusters are called CpG islands and can also be found either near or in gene promoters [4]. CpG islands in gene promoters are not methylated [4,5], therefore they allow transcription while in the rest of the genome they are heavily methylated [6]. Aberrant DNA methylation seems to happen early in endometrial tumorigenesis and it is a universal phenomenon that affects many critical genes [7].

Post translational modifications of histone proteins are another epigenetic mark. In genes where transcription is active, core histones H2A, H2B, H3 and H4 have specific acetylated lysine residues that prevent the histones from being in close contact with the DNA in the nucleosome. Thereafter, transcription factors and polymerases are free to reach coding sequences and commence transcription. Many his-tone deacetylases (HDACs) that reverse these modifications are associated with gene silencing [8]. While acetylation occurs in euchromatin, histone methylation is also an epigenetic mark popular in heterochromatin as well as in euchromatin and it is orchestrated by histone methyltransferases and histone demethylases [9]. Differential methylation in the same histone can both be a marker of gene activation or silencing [3]. Histone deacetylation that leads to gene silencing is also associated with DNA methylation, while histone marks that activate gene transcription are also observed with DNA hypomethylation.

Another mechanism that causes gene silencing is RNA-associated silencing and is induced by micro RNAs (miRNAs) [3]. miRNAs are implicated in the development, cell cycle and cell death [10]. Moreover, it has been shown in many studies that miRNAs play an important role in cancer [11]. The most interesting finding is that miRNAs can cause histone modifications, DNA methylation, changes in chromatin status [3], as well as regulation of the expression of DNMTs and HDACs which makes them key players in cancer epigenetic regulation. miRNAs have also been identified as targets of epigenetic changes.

Epigenetic modifications in type I endometrial cancer

Two different subtypes of endometrial cancer are recognized: type 1 or endometrioid (estrogen-related) and type 2 or non-endometrioid (non-estrogen related) (Figure 1). Eighty percent of newly diagnosed cases in the Western world are of type I and are mostly encountered in young or perimenopausal women under unopposed estrogenic stimulation [1]. These tumors (endometrioid carcinomas, EECs) resemble morphologically the normal endometrium and arise in a setting of endometrial hyperplasia [12]. They have usually minimal myometrial invasion, exhibit low-histological grade and are often cured with hysterectomy.

Promoter hypermethylation is the most common epigenetic mechanism found in EECs. Nieminen et al. identified 24 tumor suppressor genes whose promoters were progressively hypermethylated during the development of the disease. What precedes though this epigenetic modification and might be responsible for the appearance of it is microsatellite instability (MI). MI is present in 20-35% of EECs and it is hypothesized that it provokes alterations in many regulatory genes involved in DNA repair, apoptosis, transcriptional regulation and signal transduction that promote carcinogenesis [13]. The most common mechanism for tumor suppressor gene silencing in endometrial cancers with MI is MLH1 promoter hypermethylation and studies have shown that it is an early event in cancer progression [14]. Promoter hypermethylation is not only present in EECs with MI that lack MLH1 expression but also

Table 1. Possible epigenetic biomarkers for EEC (type 1)				
Normal cellular role	Epigenetic alteration			
DNA mismatch repair gene	Lack of expression			
Tumor suppressor gene	Gene silencing			
Tumor suppressor gene	Gene silencing			
Transcriptional factor expressed in endometrial stroma	Gene silencing			
	Normal cellular role DNA mismatch repair gene Tumor suppressor gene Tumor suppressor gene Transcriptional factor expressed	Normal cellular role Epigenetic alteration DNA mismatch repair gene Lack of expression Tumor suppressor gene Gene silencing Tumor suppressor gene Gene silencing Transcriptional factor expressed Gene silencing		

in cancer cell lines that lack the mismatch repair mechanism [15]. Furthermore, the demethylation agent 5-aza-2'-deoxycytidine of the MLH1 gene was found to initiate MLH1 expression and restore the activity of mismatch repair genes.

Besides MLH1, promoter hypermethylation has been identified in other genes such as RASS-F1A, MGMT and PTEN in tumors with MI. PTEN is a tumor suppressor gene that can be silenced through promoter hypermethylation but also with loss of heterozygosity (LOH) and mutations. PTEN is the most common mutated gene in EECs (30-50%) [16,17], and recent studies have linked PTEN promoter methylation with advanced stage in type 1 endometrial cancer [18].

MGMT is another silenced DNA repair gene that is present in 48% of EECs [19]. Loss of MGMT function leads to recognition of O(6)-methylguanine as adenine by DNA polymerases. O(6)-methylguanine is a pro-mutagenic form that leads G to A mutations [20].

RASSF1A is a human tumor suppressor gene that acts as a negative regulator of the RAS-MAPK signaling pathway, which is frequently altered in EECs. Loss of RASSF1A due to epigenetic gene silencing is correlated with increased activity of the RAS-MAPK pathway. RASSF1A silencing through promoter hypermethylation is a very common feature of advanced stage of type 1 endometrial carcinomas (74%) and is also related to higher frequency of lymph node involvement, to higher grade tumors, to higher incidence of recurrence and to lower disease-free survival [21].

HAND2 methylation has recently been detected in type 1 endometrial cancer. HAND2 encodes for a transcription factor expressed in the endometrial stroma and was found to be severely hypermethylated [22]. Premalignant endometrial lesions showed enhanced HAND2 methylation. HAND2 methylation is a good potential biomarker for EECs, however further research is required to

assess its true clinical use. The possible epigenetic biomarkers for the EECs are shown in Table 1.

Epigenetic modifications in type II endometrial cancer

Type II or non-endometrioid carcinoma (NEEC), is diagnosed in older postmenopausal women [13] and is more frequent in African-American women [23]. These tumors are not associated with estrogen and they are mostly highgrade serous or clear cell carcinomas (Figure 2). Unlike type I tumors that are mostly confined to the uterus, type II carcinomas invade deeply into the myometrium and are characterized by early extrauterine disease; as such they have to be treated in a more aggressive manner [13]. NEECs also seem to carry different genetic alterations from EECs. They are characterized by aneuploid karyotypes, LOH and aberrant p53 mutations. p53 alterations with simultaneous overexpression of the

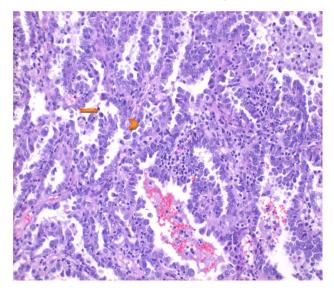


Figure 2. Serous endometrial cancer (H&E x400). The neoplastic cells are highly atypical with nuclear pleiomorphism and prominent nucleoli (arrow heads). There is cellular desquamation with many "free floating cells" (arrows).

Possible involved genes/proteins	Normal cellular role	Genetic modification
p53	Tumor suppressor gene	Inactivation
Cyclin D1/ Cyclin E	Regulators of CDK	Upregulation
Her-2/neu	Oncogene – Epidermal growth factor receptor	Upregulation
e-cadherin	Type-1 transmembrane protein	Reduction
STK15	Putative oncogene- accurate seg- regation of chromosomes during mitosis	Overexpression

Table 2. Genetic alterations in NEEC (type II)

PI3K/AKT pathway are also observed in NEECs [24]. The PI3K/AKT pathway is a signaling pathway that regulates cell cycle and promotes cell growth and proliferation. In many types of endometrial cancers it is constitutively active. Results from the above-mentioned study imply that *p*53 inactivation and activation of the PI3K/AKT pathway in high-grade endometrial carcinomas is consistent with poor prognosis.

NEECs are also associated with Cyclin D1, Cyclin E and Her-2/neu upregulation and reduced E-cadherin gene expression [1], all of the above being cellular changes that promote cell proliferation and oppose apoptosis. They also exhibit STK15 overexpression, which is responsible for increased chromosomal instability. Promoter hypermethylation seems to play a less important role in type II cancers. Promoter hypermethylation of many genes such as MLH1, PTEN, MGMT, and RASSF1A is not detected in type II tumors [25]. Loss of progesterone expression is observed in NEECs [26,27], but it is not clear yet whether its inactivation is caused by methylation, a fact that could help in designing new hormone treatment strategies for this type of cancer in the future. DNMT1 and DNMT3B are also downregulated in NEECs [28,29]. As already mentioned, DNMTs catalyze DNA methylation and a possible scenario is that this downregulation can cause global hypomethylation in NEECs and might be the reason for the histological differences between EECs and NEECs. Nevertheless, more studies need to be conducted in order to assess the different epigenetic mechanisms that underlie the different types of endometrial carcinogenesis and design prevention approaches.

Finally there is an agreement in many studies that some NEECs might arise from preexisting EECs through dedifferentiation. These tumors would possess molecular, histopathological and immunohistochemical features from both types [12]. Most of those carcinomas fall in type II category and a small subset seems to represent type 1 cancers. Possible genetic alterations found in NEEC type II are listed in Table 2.

Current research

There are many methods that have been used over the years in order to detect DNA methylation such as DNA sequencing [30], q-PCR [31], microarrays [32], mass spectrometry [33] and combined bisulfite restriction analysis (COBRA) [34]. Of these, the most popular method is sodium bisulfate treatment of DNA followed by single molecule sequencing that detects cytosine DNA methylation [35]. Throughout the years, with advances in technology, such as second generation sequencing, many more methods have been developed like whole-genome bisulfite sequencing [36,37], differential methylation hybridization analysis (DMH) [38] and deep single molecule bisulfate sequencing [39].

The most recent study was performed by Zhang et al. [40] who investigated DNA methylation in the two types of endometrial cancer as well as in normal tissues through methylated DNA immunoprecipitation sequencing (MeDIPseq) and methylation-sensitive restriction enzyme digestion sequencing (MRE-seq). The aim was to identify local differentially methylated regions (DMRs) and it is the first time that a whole-genome DNA methylation map was created for endometrial cancer. Many DMRs were common in both subtypes but some were specific to each cancer subtype and some of them were different in normal endometrium. With the use of these techniques many DMRs were identified that could not be discovered with array-based platforms [40]. These DNA methylation changes seem to be an important signature of endometrial cancer and could possibly serve as biomarkers in the future. Nevertheless, the main disadvantage of the above described techniques is that, despite their widespread use in research, they are not cost-effective and therefore they cannot be used for diagnostic routine use.

Besides DNA methylation, a number of miR-NAs was identified to play a role in the development of endometrial cancer. Many of them are involved in processes including cell death, growth, proliferation, and carcinogenesis [41]. Balch et al. [2] performed an extensive literature research and recorded all the miRNAs that are implicated in endometrial cancer. Nonetheless, a specific distinct pattern for each type of endometrial cancer has not yet been established. As miRNAs are detected in body fluids of many cancer patients, using them as cancer biomarkers is a minimal invasive way to detect the disease. Thus, investigation of these miRNAs should become top priority in endometrial cancer biology.

Conclusions

During the last years the important role of epigenetic modifications in carcinogenesis has become evident. One of the most interesting revelations was the impact of epigenetic modifications in endometrial carcinogenesis, from development till therapy. Understanding the mechanism by which epigenetic alterations along with genetic mutations, lifestyle and environmental factors lead to disease is the ultimate goal. Another important clinical impact in the use of epigenetics is in diagnosis with the help of emerging new technologies. New technologies can help in identifying the distinct methylation profile of each patient, thus leading to a more personalized treatment that would probably improve the patient's life.

References

- 1. Sorosky JI. Endometrial cancer. Obstet Gynecol 2012;120:383-397.
- Balch C, Matei DE, Huang TH, Nephew KP. Role of epigenomics in ovarian and endometrial cancers. Epigenomics 2010;2:419-447.
- Egger G, Liang G, Aparicio A, Jones PA. Epigenetics in human disease and prospects for epigenetic therapy. Nature 2004;429:457-463.
- Cross SH, Bird AP. CpG islands and genes. Curr Opin Genet Dev 1995;5:309-314.
- Herman JG, Baylin SB. Gene silencing in cancer in association with promoter hypermethylation. N Engl J Med 2003;349:2042-2054.
- 6. Bird A. DNA methylation patterns and epigenetic memory. Genes Dev 2002;16:6-21.
- 7. Jones PA, Baylin SB. The fundamental role of epigenetic events in cancer. Nat Rev Genet 2002;3:415-428.
- 8. Jones PA, Baylin SB. The epigenomics of cancer. Cell 2007;128:683-692.
- 9. Cedar H, Bergman Y. Linking DNA methylation and histone modification: patterns and paradigms. Nat Rev Genet 2009;10:295-304.
- 10. Lujambio A, Calin GA, Villanueva A et al. A microRNA DNA methylation signature for human cancer metastasis. Proc Natl Acad Sci U S A 2008;105:13556-13561.
- 11. Weber B, Stresemann C, Brueckner B, Lyko F. Methylation of human microRNA genes in normal and neoplastic cells. Cell Cycle 2007;6:1001-1005.

- 12. Matias-Guiu X, Catasus L et al. Molecular pathology of endometrial hyperplasia and carcinoma. Hum Pathol 2001;32:569-577.
- 13. Catasus L, Prat J. Molecular genetics of endometrial carcinoma. Diagn Histopathol 2009;15:554-563.
- Esteller M, Catasus L, Matias et al. hMLH1 promoter hypermethylation is an early event in human endometrial tumorigenesis. Am J Pathol 1999;155:1767-1772.
- 15. Catasus L, Matias-Guiu X, Machin P et al. Frameshift mutations at coding mononucleotide repeat microsatellites in endometrial carcinoma with microsatellite instability. Cancer 2000;88:2290-2297.
- Salvesen HB, MacDonald N, Ryan A et al. PTEN methylation is associated with advanced stage and microsatellite instability in endometrial carcinoma. Int J Cancer 2001;91:22-26.
- Risinger JI, Hayes AK, Berchuck A, Barrett JC. PTEN/ MMAC1 mutations in endometrial cancers. Cancer Res 1997;57:4736-4738.
- Hayes MP, Wang H, Espinal-Witter R et al. PIK3CA and PTEN mutations in uterine endometrioid carcinoma and complex atypical hyperplasia. Clin Cancer Res 2006;12:5932-5935.
- Vassileva V, Millar A, Briollais L, Chapman W, Bapat B. Genes involved in DNA repair are mutational targets in endometrial cancers with microsatellite instability. Cancer Res 2002;62:4095-4099.
- 20. Esteller M. Epigenetic lesions causing genetic lesions

in human cancer: promoter hypermethylation of DNA repair genes. Eur J Cancer 2000;36:2294-2300.

- 21. Pallares J, Velasco A, Eritja N et al. Promoter hypermethylation and reduced expression of RASSF1A are frequent molecular alterations of endometrial carcinoma. Mod Pathol 2008;21:691-699.
- 22. Jones A, Teschendorff AE, Li Q et al. Role of DNA methylation and epigenetic silencing of HAND2 in endometrial cancer development. PLoS Med 2013;10:e1001551.
- 23. Allard JE, Maxwell GL. Race disparities between black and white women in the incidence, treatment, and prognosis of endometrial cancer. Cancer Control 2009;16:53-56.
- 24. Catasus L, Gallardo A, Cuatrecasas M, Prat J. Concomitant PI3K-AKT and p53 alterations in endometrial carcinomas are associated with poor prognosis. Mod Pathol 2009;22:522-529.
- 25. Risinger JI, Maxwell GL, Berchuck A, Barrett JC. Promoter hypermethylation as an epigenetic component in Type I and Type II endometrial cancers. Ann N Y Acad Sci 2003;983:208-212.
- 26. Bonfitto VL, de Angelo Andrade LA. p53, estrogen and progesterone receptors in diagnostic curettage for endometrial adenocarcinoma and their correlation with morphological data and disease stage at hysterectomy. Sao Paulo Med J 2003;121:163-166.
- 27. Lax SF, Pizer ES, Ronnett BM, Kurman RJ. Clear cell carcinoma of the endometrium is characterized by a distinctive profile of p53, Ki-67, estrogen, and progesterone receptor expression. Hum Pathol 1998;29:551-558.
- Liao X, Siu MK, Chan KY et al. Hypermethylation of RAS effector related genes and DNA methyltransferase 1 expression in endometrial carcinogenesis. Int J Cancer 2008;123:296-302.
- 29. Reid-Nicholson M, Iyengar P, Hummer AJ, Linkov I, Asher M, Soslow RA. Immunophenotypic diversity of endometrial adenocarcinomas: implications for differential diagnosis. Mod Pathol 2006;19:1091-1100.
- 30. Frommer M, McDonald LE, Millar DS et al. A genomic

sequencing protocol that yields a positive display of 5-methylcytosine residues in individual DNA strands. Proc Natl Acad Sci U S A 1992;89:1827-1831.

- 31. Eads CA, Danenberg KD, Kawakami K et al. MethyLight: a high-throughput assay to measure DNA methylation. Nucleic Acids Res 2000;28:E32.
- 32. Ushijima T. Detection and interpretation of altered methylation patterns in cancer cells. Nat Rev Cancer 2005;5:223-231.
- 33. Ehrich M, Nelson MR, Stanssens P et al. Quantitative high-throughput analysis of DNA methylation patterns by base-specific cleavage and mass spectrometry. Proc Natl Acad Sci U S A 2005;102:15785-15790.
- 34. Xiong Z, Laird PW. COBRA: a sensitive and quantitative DNA methylation assay. Nucleic Acids Res 1997;25:2532-2534.
- Varley KE, Mitra RD. Bisulfite Patch PCR enables multiplexed sequencing of promoter methylation across cancer samples. Genome Res 2010;20:1279-1287.
- Cokus SJ, Feng S, Zhang X et al. Shotgun bisulphite sequencing of the Arabidopsis genome reveals DNA methylation patterning. Nature 2008;452:215-219.
- 37. Lister R, Pelizzola M, Dowen RH et al. Human DNA methylomes at base resolution show widespread epigenomic differences. Nature 2009;462:315-322.
- Yan PS, Potter D, Deatherage DE, Huang TH, Lin S. Differential methylation hybridization: profiling DNA methylation with a high-density CpG island microarray. Methods Mol Biol 2009;507:89-106.
- Varley KE, Mutch DG, Edmonston TB, Goodfellow PJ, Mitra RD. Intra-tumor heterogeneity of MLH1 promoter methylation revealed by deep single molecule bisulfite sequencing. Nucleic Acids Res 2009;37:4603-4612.
- Zhang B, Xing X, Li J et al. Comparative DNA methylome analysis of endometrial carcinoma reveals complex and distinct deregulation of cancer promoters and enhancers. BMC Genomics 2014;15:868.
- 41. Boren T, Xiong Y, Hakam A et al. MicroRNAs and their target messenger RNAs associated with endometrial carcinogenesis. Gynecol Oncol 2008;110:206-215.